After a year in which not one month could boast of “normal” weather, the 2011 fall mushroom season was wonderfully surprising, producing some results quite different from typical years. After receiving more snow in the months of April and May than average for the entire winter in the North Cascades, followed by a wet and very cold June, the mushrooms in our area remained about 6 to 8 weeks behind schedule for the duration of the season. Some species, such as *Boletus edulis* delayed or missed fruiting in areas, like the Scott Paul Trail, where they normally thrive. This could be said of nearly all of the alpine species, and many of the mycorrhizal mushrooms throughout our area, even in the lowland forests. On all my fall forays, I kept coming up empty handed. Where were all the mushrooms? By the first week of October, with the show just days away, I started to panic. In fact, I called Fred Rhoades and suggested that we get together and compile a passable photo array to appease the public’s desire to see something fungal at the Annual Fall Mushroom Show. Fred advised that we not be so hasty, that a lot could happen in the space of a week. Fred was correct. Fortune smiled upon us, and the autumn rains arrived, giving us three solid days of precipitation, when we, and the mushrooms, needed it the most.

We ended up not only with far more diversity than I expected, but finished with 42 more species of fungi than we have ever shown before! While it remained a very sparse year for the mycorrhizal mushrooms, the saprophytes rushed in to save the day. The woods were chock full of wood-loving decomposers, some quite rare, and some on display for the first time at our fall show.

Curiously, the chanterelles seemed to be oblivious to the odd conditions; species in the genus *Cantherellus* cropped up in very respectable numbers, and in some cases, outright abundance.

The late arrival of a myriad of mushrooms was by no means the only element of the amazing success of the 2011 Fall Exhibit. The volunteer force of collectors, tray arrangers,
labelers, kitchen staff, and identifiers, performed impeccably to coordinate and put together an awesome set of displays by the time the doors opened at noon on Sunday to an enthusiastic throng of mycophiles. More people stepped up to help collect for the show than any other, an impressive effort in a very challenging year. Because of those volunteers, this year’s show featured the largest collection of mushrooms ever assembled by the Northwest Mushroomers Association. As the chairperson of the show, I offer my congratulations and heartfelt thanks to all who worked so diligently to bring it all together. A special thanks to Dr. Fred Rhoades, our invaluable science advisor, Buck McAdoo, Christine Roberts, Erin Moore, and Larry Baxter, our team of ace identifiers, and special mention of thanks to Margaret Dilly, who, in addition to being an identifier with the rest of the identification team, coordinates all of the fine details of assigning people to tray staging, and is responsible for supervising the arrangement of trays on the display tables, a monumental undertaking. Very honorable mention to Chuck Nafziger for his work on the centerpiece, which is better each year, and his diligence on identifying, labeling, and staging of the plethora of polypores that made into the show, and finally, to Sonya, for traversing all the way to the Olympic Peninsula to bring back some of the most interesting fungi in the show.

I’m very glad Fred talked me out of the photo array!
Mushroom of the Month: *Floccularia albolanaripes* (Atkinson) Redhead  
By Buck McAdoo

One of the most colorful surprises of the October 2011 Fall Show was the enigmatic appearance of *Floccularia albolanaripes*. The fleshy yellow fruiting bodies dominated the *Armillaria* section, the genus where it had been before. Also known as ‘The Shaggy Stalk Mushroom’ if you follow Mary Wells, or ‘The Scaly Bracelet’ according to the McK-nights, this was the first time it had arrived at our fall show. Since there are only seven *Floccularias* in the world, it made its presence even more special. The problem with the species at the fall shows is that so many people bring mushrooms in that we often can’t match mushroom with finder. So, whoever you are, congratulations on landing ‘the mushroom of the month’.

But why *Floccularia*? George Atkinson placed it in *Armillaria* in 1908 from a collection found by E.R. Lake from Corvalis, Oregon on November 6, 1906. There it lived for 81 years. Then in 1957, Pouzar erected the genus *Floccularia*. This was created for those species of *Armillaria* with no black rhizomorphs, fleshy fruiting bodies with some yellow in them, cap margins with appendiculate velar shards, and smooth, amyloid spores. But *Floccularia albolanaripes* is not an European species, and it took time before Dr. Redhead herded it into its proper genus in 1987.

As for identification, I had seen this species only once before. This was back at the Baker Lake foray in October of 2000. The caps had been brilliant yellow, glabrous, and viscid. This is how they often start out. The specimens seen here were merely sticky and had flattened scales and fibrils of a cinnamon-brown color. The literature tells us that this is how the species changes in appearance as it ages. Older caps can turn from yellowish to brownish. Its closest relative, *Floccularia luteovirens*, differs by having crowded, erect scales on both cap and stem. A good comparison of the two species can be seen on adjacent pages in Alexander Smith’s A Field Guide to Western Mushrooms.

Caps of *Floccularia albolanaripes* are 5-15 cm wide, broadly convex and usually umbonate. They are viscid at first, then sticky as they dry. The color is bright yellow becoming pallid at the margins, and usually decorated with flattened darker scales and fibrils. Cap margins are at first inrolled and covered with the white, appendiculate veil remnants. The context is white, but yellow beneath the cuticle. According to Wells and Mitchel, the caps are sensitive to light. In Colorado they can bleach to a cream color in sunlight or be cinnamon-brown in the shade. The gills are adnexed to notched, close to subdistant, and can have straight or serrated edges. They are white at first, then yellowish in age. The stems are 3-8 cm long and 1-2 ½ cm thick. The apex is smooth and white to pale yellow. Then there is an abrupt change. The rest of the stem is sheathed in a belt of dense white to yellowish cottony scales with brownish tips. The top of this sheath is a ring of the white cottony velar remnants that separated from the cap margin. A gorgeous photo of this can be viewed at Mykoweb.com. Atkinson reported the stems to be hollow. The odor and taste are reported as mild by all authors except for Wells & Mitchel, who found the taste to be sour and acrid when raw.

Along the west coast, *F. albolanaripes* is found mostly with alder and oak from fall through winter down into California. David Biek found it with yellow pine and oak through spring in Northern California. Wells & Mitchel reported it as common in high aspen meadows in Colorado in the summers. Barrows found it in New Mexico under mountain conifers. Dr. Dennis Thurber found it at Aspen, Colorado with Engelmann spruce. And Jack States thought so much of it that he put it on the cover of his popular guide, Mushrooms and Truffles of the Southwest. Calvin Kauffman found it in the Olympics in 1922, but disagreed with Atkinson’s spore sizes. (Just
for the record, we looked at the spores and came up with 3.9-5.2 x 5.4-7.4 microns, which were closer to Atkinson’s.

In 1976, Smith & Mitchel found an albino form, which they introduced as *Armillaria albolanaripes* form alba. It is reputed to be one of the world’s most beautiful mushrooms. As for the typical form, you can’t confuse it with much. *Floccularia pitkinensis* has fleshy grayish caps with tinges of yellow. *Amanita franchettii* differs by having free gills, yellow warts on the cap, and loose yellow velar material on the stem. Jack States felt that they looked like large yellow *Pholiotas*, except for the habit of growing on the ground and the white spores instead of rusty ones. Otherwise, *Floccularia albolanaripes* seems a fairly safe mushroom for beginners.

And this brings up edibility. The Shaggy Bracelet has long been considered inferior to its floccose cousin, *Floccularia luteovirens*, highly esteemed in Europe and the Rockies. Here now is a sampling of the opinions from popular guides:

Wells & Mitchel – ‘Edibility unknown’.
David Biek – ‘Of little value as an edible’.
Jack States – ‘Edible but lacks the quality of *A. straminea*’, (an earlier name for *F. luteovirens*).
David Arora – ‘Edible but insipid’.
Helene Schalkwijk-Barendsen – ‘The taste is bland’.
McKenny, Stuntz, & Ammirati – ‘Of uncertain edibility’.
Mike Woods of Mykoweb – ‘Edible and excellent. A greatly under-appreciated mushroom’.

Well, as far as I’m concerned, Woods has got it right. I took three of the specimens from the show down to my boat galley. Simply sautéed in butter, they were among the ten best mushrooms I’ve ever tasted. Besides having just the right ratio of crunchy exterior and juicy interior, the flavor had a gourmet aftertaste that brought up visions of manna. I can only attribute its lukewarm reception to circumstance. *Cortinarius caperatus* comes to mind. I have eaten it four times and only once considered it choice. The other three samplings were insipid to poor. Maybe both of these species have to be in prime condition and with the proper substrate in order to taste good.

For those increasing members in our club who enjoy microscopic characters, we discovered that *Floccularia albolanaripes* had no cystidia whatsoever. It had parallel gill trama with hyphae 5-11.4 microns wide. The clavate basidia were 4-spored and measured 6.8-7.8 x 34-36 microns. Spores were smooth and ellipsoid. The pileipellis consisted of radially repent hyphae measuring 6-12 microns wide. A.H. Smith noted occasional clamps and a particularly narrow subhymenium in the gill trama. He and Mitchel also noted that the central strand in the gill trama tended to be more inflated than the surrounding hyphae. We didn’t see this phenomenon, but it doesn’t mean it couldn’t happen.

Meanwhile, keep your eyes open. Although possibly a first for Whatcom County, more could follow. If you don’t want to risk eating them, bring them to one of our experts. We’ll exert a tax or consume the whole collection if you prefer.

Bibliography
2011 Fall Show Observations and Report

By Buck McAdoo

It’s always a wild card on how the annual weather patterns impact the mushrooms. This year was rather untypical in that the snow pack remained at high elevations all through the summer months. I doubt if Artist’s Point ever did open. One of the reasons for this was our unusually cool, wet summer. It seemed to rain off and on all through July and into August, with summer finally arriving in September. Since fungi require moisture, I speculated that this would be the most bountiful fall we ever had. When Steve Trudell advised us in September that this would not be a good mushroom year, I couldn’t fathom the connection.

As the fall progressed with no reports of Boletus edulis, and just occasional sightings of our ubiquitous chanterelles, Steve’s prediction seemed more accurate than not. The final blow came just a week before our show. Fred Rhoades organized a foray at Silver Lake. Half the forayers stayed around Silver Lake. The rest of us drove off to one of my favorite collecting sites up the Hannegan Pass Rd. Except for one magnificent fruiting of the earth star, Geastrum saccatum, there was nothing else there. Just acres of moist, undulating moss with not even an Inocybe to break the green. The people at Silver Lake fared a lot better, but it was still way below normal for this time of year. Our Fall Show chairman was even heard to be muttering about canceling the show. There was the suggestion that we could have photos of mushrooms on all the tables. Another idea was to fill the tables with Gymnopus peronatus. Maybe the public could be appeased by the quantity and not notice so much the lack of diversity.

Then came the show. To me, anyway, it was like the New York Mets winning their first World Series. Nature somehow came through. Lo and behold, boxes of unusual things began arriving at Bloedel-Donovan from all over the state. Vince had gone all the way out to Easy Pass on Highway 20. Sonia had covered the Olympic Peninsula. Fred had stopped by his favorite sites on the way back from Spokane. Jairul brought a desiccated clump of the blue-black chanterelle from the show at Everett the week before. These are just the endeavors we knew about. But the entire club must have gone to similar lengths because from seemingly nothing, we broke the club record for different species at the fall show! At one point I heard the number as 297 species. Since then, Fred has adjusted the list to eliminate redundancies and perhaps add a few species that had lost their identification cards.

Each season is totally different. Two years ago, Russula boxes almost extended all the way across the room. Last year, Cortinarius ruled. But this year was the year of the Pholiota. There seemed to be more boxes of Pholiota than
anything else, an odd thing for a genus that usually peaks here in November, especially considering that the accompanying fungi were mostly composed of species that show up here in September. Lisa McAvoy and her team of helpers did a bang-up job of getting these Pholiotas on the table. We even saw Pholiota albocrenulata for the first time.

But everyone involved deserves praise. Folks I hardly knew were scurrying around identifying things at a rate I hadn’t seen before. As a club, I believe we have turned a corner. At some point around mid-afternoon, I drifted around the tables looking for obvious misidentifications. There really weren’t any! Just in case I missed something, I asked new member Kevin Bi, a keenly perceptive connoisseur from White Rock, to make the rounds also. He questioned Suillus lakei and Lepiota brebissonii. I would have questioned the latter also if Steve Trudell hadn’t set us straight in September. Suillus lakei and Suillus cavipes only differ visually in the presence of a hollow stem in the latter. When the stems turned out to be solid, it looked like we had done our homework. I seriously doubt whether any other club of our size in America has as many good identifiers.

A lot of the credit has to go to Fred and Margaret. Both have taught mushroom identification classes for years. They both deserve the annual NAMA award for contributions to mycology. For this fall show, they both worked hard on streamlining the identification process ahead of time. There were new lists of fungi showing what we already had labels for. Species names in blue indicated there was a more modern name for that species that could be looked up in another list. For the larger genera, the names of species in their boxes were all in alphabetical order. This is a huge factor when you are pressed for time. There were, of course, some minor snafus. The labels for the Birds’ Nest fungi couldn’t be found at first. A few specimens brought to the show were reported as lost. This is always agonizing for the person who brought them. Sometimes they are found again, and sometimes not. It’s become part of the mystique of the fungi.

All in all, this year’s show represented an amazing effort by both those who went out and found the mushrooms and those who identified them later. You all know who you are. There are now just too many of you to list you individually. We can now reserve that list for the fungi.

2011 Lummi Island Foray: My First Host

By Richard Mollet

Nope, I’m not referring to white wafers but rather my first experience hosting a foray (10/22/11). I awoke at 7:00 AM to a pouring rain. Hoping that the weather would improve I showered, ate breakfast, dressed and by 8:30 the pour had increased to deluge status. Adopting an “it is what it is what it is” attitude I set out wondering if anyone would show up at the Gooseberry Point rendezvous (one never knows about shroomers). As I turned onto Haxton by Casino Corner the rain started to let up and by 9:20 the sun began to shine.

At 9:45 witnessed the arrival of nine other intrepid club shroomers and off we went to the Otto Preserve on Lummi Island. We had no sooner arrived than we were joined by about 15 islanders, who unbeknownst to me had been invited to share our foray. I explained that we were not elitists and they were welcome to share whatever we had.

Groups spread out over the vast acreage while I set up the “kitchen” and Buck set up an identification area in the lodge. I then took a short walk outside and soon espied my first blewit (Lepista personata). I’d seen photos and examples at our fall show but never found one in situ.

After about two hours, seekers started to return with
quite a plethora of mushrooms, easily at 50-60 types. Everyone was feeling good with what they found and some were looking forward to a tasty supper at home. Speaking of which, the potluck lunch was quite interesting itself.

We remembered to save our identifying labels as someone had indicated that Fred Rhoades wished to compile a catalogue of Lummi Island mushrooms. Just as I closed the trunk lid of my car it started to rain again. It is what it is.

**Bowman Bay Field Trip November 5, 2011**

*By Margaret Dilly*

On a beautiful fall morning in November we arrived at Bowman Bay State Park to find Maggie Sullivan our faithful member had already set out the signs and was hauling the club supplies up to the shelter. This area was closed for the winter so toilet facilities were a bit of a walk away but as opposed to last year we did have running water at the site. Soon with the help of a few other early arrivals who had already found some nice specimens, tables were moved around and set ready for the day. Claude made a nice roaring fire and the day began with coffee and cookies. Soon, baskets were in hand and the eager hunters dispersed in all directions. The hunt began.

Soon Larry Baxter and Kay Kelly arrived from Camano Island. Larry spent the day at the ID table with me, sorting and identifying mushrooms. More members and friends arrived as the day went on, among them who were able to help with the identification, were Dick Morrison, Harold Mead, Evan Sanford and Chuck Nafziger. The collections varied from a tiny *Strobilluris tullisatus* and a beautiful collection of *Phaeolepiota aurea*. As impressive as they are, eating them is not recommended. Just ask Jack. New and old members alike were very helpful throughout the day with the display and showed great interest in learning more about ID. The fall identification class should fill quickly. We didn’t have time to identify all species but of those we did came to 66 (listed in the Species Annex, at the back of this issue). This is good for this year and the short time we had to hunt.

Thirty three people signed in and shared a wonderful potluck lunch. As always Fien was at the stove cooking up hot goodies for all to savor. Thanks to one of our generous members, chanterelles (*Cantharellus formosus*) were donat-ed, cooked up and enjoyed by all. No Blewitts (*Lepista nuda*) were found. Traditionally this area produces them in abundance every year, but not one appeared this year. The strange weather conditions we have seen have definitely affected the fungal fruiting this year. It began to cool by early afternoon and the clean up began.
Hunters gathered up their baskets with a few specimens they wanted to keep and headed for home. Maggie and Harold and several loyal members helped with clean up. All in all it was a successful fall outing. Now we can all look forward to the new mushroom year starting with the Survivors Banquet, March 3rd, and Morel Madness in May.

Be thinking of volunteering to help for both of these events. I look forward to seeing all of you then.

2012 NMA MONTHLY MEETING SPEAKERS

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*Look for much more information on David Arora’s visit in the next issue of MushRumors

“SPONGEBOB” MUSHROOM NAMED

Truth Stranger Than Fiction?

Christine Dell’Amore

National Geographic News, June 16, 2011

The new species, Spongiforma squarepantsii—found in 2010 in Sarawak, Malaysia—has a spongy appearance that reminded scientists of TV’s Spongebob Squarepants.

“It’s just like a sponge with these big hollow holes,” San Francisco State University’s Dennis Desjardin said in a statement. “When it’s wet and moist and fresh, you can wring water out of it and it will spring back to its original size. Most mushrooms don’t do that.” There’s only one other species known so far in the Spongiforma genus; it lives in central Thailand and has a different color and odor. S. squarepantsii has a bright orange hue and smells “vaguely fruity or strongly musty,” according to the study, published in May in the journal Mycologia. When Desjardin and colleagues looked at the new mushroom under a scanning electron microscope, they
found even more spongy similarities—for instance, the spore-producing area of the fungus resembles a seafloor carpeted in tube sponges. One thing’s for certain—there are more, more weird fungi out there. Only five percent of Earth’s fungi species have been found, and there may be up to three million still unknown.

**Silver Lake Foray October 1, 2011**

Agaricus albolutescens
Agrocybe praecox
Agrocybe sp.
Albatrellus flettii
Amanita fulva
Bolbitius vitellinus
Boletus chrysenteron
Boletus zelleri
Cantharellus formosus
Clitocybe connata
Coltricia perennis
Conocybe tenera
Coriolus versicolor
Coprinopsis lagopus group
Coprinellus micaceus
Crepidotus applanatus
Cystoderma terreii
Fomes fomentarius
Fomitopsis pinicola
Ganoderma applanatum
Geastrum saccatum
Geastrum triplex
Gomphus floccosus
Gymnopilus ventricosus
Gymnopus confluens
Gymnopus peronatus
Hypomyces lactifluorum
Inocybe calamistrata
Inocybe sororia
Jahnoporus hirtus
Lactarius obscuratus
Lactarius scrobiculatus
Leccinum scabrum
Mycena aurantiidisca
Mycena filopes
Mycena galericulata
Nidula candida

Panaeolina foenisecii
Paxillus involutus
Peziza repanda
Phaeolus schweinitzii
Pholiota flammans
Pholiota squarrosa
Pleurotus elongatipes
Pluteus cervinus
Pluteus lutescens?
Polyporus badius
Polyporus melanopus
Psathyrella torpens?
Ramaria stricta
Russula sp.
Russula brevipes
Russula brunneola
Russula dissimilans (formerly R. nigricans)
Russula isabelliniceps
Russula occidentalis
Russula placita?
Russula xerampelina
Scleroderma fuscum
Sparassis crispa
Strobilurus trullatus
Tapinella atrotomentosa
Thelephora palmata
Tricholomopsis decora

Lichens
Lobaria oregana
Peltigera canina
Peltigera neopolydactyla
Lummi Island Foray Species List

The species list for the October 22, 2011, foray at the Otto Preserve consisted of 80 species, not all readily identifiable. This preserve is turning out to be such a good fungal site that it might be worth while for the preserve to invest in a copy of Mushrooms Demystified for detailed referencing. While the vast majority of species found on this foray were common to the mainland, a few odd things occurred that we hadn’t seen before. I’ll bring those up after the list.

The List
Agaricus hondensis
Lepiota decorata
Agaricus ‘moelleri’
Lepiota sequoiarum
Armillaria nabsona
Lepista nuda
Armillaria solidipes
Leucoagaricus leucothites
Arrhenia sp.
Leucoagaricus rubrotinctoides
Ascocoryne ‘sarcoides’
Leucopaxillus amarus
Boletus chrysenteron
Lycoperdon perlatum
Cantharellus formosus
Marasmiellus candidus
Cheimonophyllum candidissimum
Melanoleuca melaleuca
Chlorophyllum olivieri
Mycena adonis
Clitocybe connata
Mycena alcalina group
Clitocybe deceptiva
Mycena citrinomarginata
Clitocybe inversa
Mycena pura
Clitocybe sp.
Nolanea hebes
Clitocybula atrialba
Panaeolina foenisecci
Clitopilus prunulus
Paxillus involutus
Coprinus comatus
Phaeolus schweinitzii
Cortinarius mucosus group
Phlebia radiata

By Buck McAdoo

Crepidotus sp.
Phlebia tremellosa
Cystoderma terreii
Pleurotus longinquus
Fomitopsis cajanderi
Pluteus cervinus
Fomitopsis pinicola
Polyporus varius
Galerina sp.
Pseudohydnum gelatinosum
Galerina sp.
Rickenella fibula
Geastrum saccatum
Russula fragilis
Geastrum triplex
Russula murrillii
Gomphidius subroseus
Russula vernonia
Gymnopilus sapineus group
Russula xerampelina
Gymnopus dryophilus
Spathularia flavida
Gymnoporus peronatus
Stereum gausapatum
Hebeloma crustuliniforme
Stereum hirsutum
Heterobasidion annosum
Stribilurus occidentalis
Hygrophoropsis aurantiacus
Stribilurus trullisatus
Hypholoma fasciculare
Stribpharia ambiguum
Hypomyces chrysospermus
Suillus caerulescens
Hypomyces lactifluorum
Suillus lakei
Lactarius luculentus var. laetus
Tapinella atrotomentosa
Lactarius obscuratus
Trametes versicolor
Lepiota cristata
Xeromphalina campanella
Xeromphalina cornui
Xylaria hypoxylon

continued on page 11
Sometimes you will see the word ‘group’ after a species name. This generally means the species is part of a complex of close relatives or look-alikes and would require both microscopic and DNA analysis to get to the correct name. The name is likely to be the correct name in the popular guides and maybe sufficient for the purposes of the Otto Preserve. If you a species name in parenthesis, it means there could be some question over it, or a name change is in the works. In the case of Agaricus ‘moelleri’ in the list above, the name ‘moelleri’ belongs to a species from Europe, and ours will soon be getting a new name since it differs markedly in DNA profiling. In the case of Ascocoryne ‘sarcoides’, a little purple subgelatinous species on wood, the fruiting bodies were so distorted that they did not resemble the cupular shapes normally associated with the species. Sadly, the specimen was lost on the identification table.

One of the most interesting species was found by Richard Mollette, our foray host. This was a flabby and fragile grayish Clitocybe in the lawn not far from the front door. It might have been the morbid Clitocybe, Clitocybe morbifera, but I didn’t collect it for study because I had left my camera behind due to heavy rain at the time of departure. Ditto, for the lawn Galerinas. One looked like photos of Galerina hypnorum, the other Galerina graminea. But you can’t name the species in this genus without microscopic verification since so many of them look alike.

Lummi Island harbors interesting fungi. The only Limacella (a genus that looks like Amanita with a slimy cap surface) I’ve ever seen in the state came from this island. It was Limacella roseicremea, a cream colored species with a pinkish tinge. Nancy Burnette brought it to one of our club meetings back in the day.

**FUNGUS SPREADS SOUTH FROM B.C., BECOMES MORE DEADLY**  
*various sources*

(Featured in PSMS Sporeprints, June 2010)

It sounds like a villain from a science fiction film. Cryptococcus gattii—an airborne fungus that appeared on Vancouver Island in the late 1990s—is real, and it’s gathering strength as it spreads to the south. According to research published April 22 in the journal Public Library of Science Pathogens, it has mutated into a more lethal strain since it moved into Oregon. The fungus has now spread to California.

So far, Washington cases of cryptococcosis caused by C. gattii have been reported in Whatcom, King, and San Juan counties. “The cases in Washington—and there’s only been eight or nine of them—are strongly linked to the British Columbia strain,” said Dr. Tom Locke, public health officer for Clallam and Jefferson counties.

Fortunately, though potentially deadly to humans and animals infections of C. gattii are still rare. According to the Oregon Department of Human Services (April 26, 2010), since 2004 about 50 people have been identified with the illness in Washington, Oregon, and California and about 10 people have died. Cryptococcus gattii is not transmitted from person to person or carried by insects or animals. Rather, the fungus forms spores that are blown in the wind or moved by disturbances of the soil. People who stir up the soil—landscapers, loggers, outdoor recreationalists—are the most likely to encounter the fungus. Besides being spread by the wind, the fungus can be spread by humans on shoes and even on car tires.

The spores are inhaled and colonize the lungs before they spread throughout the body. Symptoms include shortness of breath, chest pain, long-lasting coughs, fever, and headaches—even weeks after exposure. Most cases are like a pneumonia that slowly gets worse and worse. Treatment involves six to eight weeks of intravenous antifungal medications followed by months of pills.
2011 Fall Show Species List  Compiled by Fred Rhoades and margaret Dilly

Now 291 (179 gilled, 96 nongilled, 12 lichens, 4 slime molds)

Ascomycota

Aleuria aurantia "Orange fairy-cup"
Bisporella citrina
Chlorencoelia versiformis
Chlorociboria aeruginascens "Green stain cup"
Gyromitra infula "Hooded false morel"
Helvella solitaria
Hymenoscyphus sp.
Hypomyces chrysospermus
Hypomyces lactifluorum "Lobster mushroom"
Otidea onotica
Peziza arvernensis
Tarzetta cupularis
Xylaria hypoxylon "Candlesnuff fungus"

Gasteromycetes (Puffballs, etc.)
Bovista plumbea
Crucibulum laeve
Cyathus stercoreus
Cyathus striatus
Geastrum saccatum
Geastrum triplex
Lycoperdon (Morganella) pyriforme
Lycoperdon nigrescens (L. foetidum)
Lycoperdon perlatum
Lycoperdon umbrinum
Scleroderma bovista
Scleroderma cepa
Truncocolumella citrina
Vascellum lloydianum (V. pratense)

Jelly Fungi
Calocera cornea
Dacrymyces chrysospermus (D. palmatus)
Pseudohydnum gelatinosum
Tremella mesenterica
Tremiscus (Phlogiotis) helvelloides

Boletes

Boletus chrysenteron
Boletus edulis "King bolete"
Boletus fibrillosus
Boletus mirabilis "Admirable bolete"
Boletus zelleri
Chalciporus (Boletus) piperatus
Leccinum holopus
Leccinum scabrum "Birch bolete"
Phylloporus rhodoxanthus "Gilled bolete"
Suillus caerulescens
Suillus lakei
Suillus luteus "Slippery jack"

Poly pores, crusts & the like
Antrodia serialis
Bondarzewia mesenterica (B. montana)
Cerrena (Daedalea) unicolor
Chondrostereum (Stereum) purpureum
Coltricia cinnamomea
Coltricia perennis
Daedaleopsis confragosa
Fomes fomentarius
Fomitopsis (Fomes) cajanderi
Fomitopsis officinalis
Fomitopsis pinicola
Ganoderma applanatum
Ganoderma oregonense (G. tsugae)
Heterobasidion (Fomes) annosum
Janoporus (Polyporus) hirtus
Laetiporus conifericola (L. sulphureus)
Lenzites betulina
Oligoporus leucospongia
Phaeolus schweinitzii
Phlebia tremellosa
Piptoporus betulinus
Polyporus melanopus
Pycnoporellus alboluteus
Pycnoporus cinnabarinus
Stereum hirsutum
Thelephora terrestris
Trametes (Coriolus) versicolor
Trametes (Coriolus) hirsuta
Tyromyces chioneus

Teeth fungi
Hericium abietis "Coral Hydnum"
Hydnellum peckii
Hydnellum regium
Hydnum (Dentinum) repandum "Hedgehog mushroom"

Corals
Artomyces (Clavicorona) pyxidatus
Clavaria vermicularis
Clavariadelphs truncatus
Clavulina cinerea
Clavulina cristata
Clavulinopsis laeticolor
Ramaria abietina
Ramaria acriccesiscens
Ramaria araisopora
Ramaria cystidiophora var. fabiolens
Ramaria sandaracina var. euosma
Sparassis crispa "Cauliflower fungus"

Chanterelles
Cantharellus formosus (C. cibarius) "Pacific golden chanterelle"
Cantharellus subalbidus "White chanterelle"
Craterellus (Cantharellus) tubaeformis "Winter chanterelle"
Gomphus bonarii
Gomphus clavatus "Pig's ears"
Gomphus kaufmanii
Polyozellus multiplex "Blue chanterelle"

Agarics (gilled)
Agaricus augustus "The prince"
Agaricus bitorquis (A. rodmani)
Agaricus campestris "Meadow mushroom"
Agaricus cupreobrunneus
Agaricus integer
Agaricus moelleri (A. praeclaresquamosus)
Agaricus nivescens
Agaricus silvaticus
Agrocybe paludosa
Amanita aprica
Amanita gemmata
Amanita muscaria "Fly Amanita"
Amanita smithiana (A. solitaria)
Ampulloclitocybe (Clitocybe) avellaneoalba
Ampulloclitocybe (Clitocybe) clavipes
Armillaria solidipes (A. ostoyae, A. mellea) "Honey mushroom"
Asterophora parasitica
Cantharellula (Clitocybe) umbonata
Catathelasma ventricosum
Cheimonophyllum (Pleurotus) candissimum
Chlorophyllum (Lepiota) olivieri
Chlorophyllum (Lepiota) rachodes "Shaggy parasol"
Chroogomphus tomentosus "False wooly chanterelle"
Chrysomphalina (Omphalina) chrysophylla
Chrysomphalina aurantiaca (Omphalina luteicolor)
Clitocybe connata (C. dilatata)
Clitocybe dealbata "Sweat-producing Clitocybe"
Clitocybe deceptiva
Clitocybe diatreta
Clitocybe odora
Clitocybe sinopica
Clitocybula (Clitocybe) atrialba
Clitopilus prunulus "Sweetbread mushroom"
Collybia cirrhata
Coprinellus (Coprinus) micaceus "Mica caps"
Coprinopsis atramentaria "Inky caps"
Coprinus comatus "Shaggy mane"
Cortinarius (Rozites) caperatus "Gypsy mushroom"
Cortinarius acutus
Cortinarius alboviolaceus
Cortinarius anomalus
Cortinarius vanduzerensis
Cortinarius violaceus "Violet Cortinarius"
Cortinarius sp. (green corn)
Crepidotus applanatus
Crepidotus epibryus (C. herbarum)
Cystodermella (Cystoderma) cinnabarina (Cystoderma terreyi)
Entoloma rhodopolium
Floccularia (Armillaria) luteovirens
Gomphidius subroseus
Gomphidius glutinosus
Gomphidius smithii
Gymnopilus bellulus
Gymnopilus croceoluteus
Gymnopilus penetrans
Gymnopilus punctifolius
Gymnopilus spectabilis
Gymnopus (Collybia) acervatus
Gymnopus (Collybia) confluens
Gymnopus (Collybia) peronatus
Hebeloma incarnatum (H. crustuliniforme)
Hebeloma mesaphaeum
Hebeloma sacchariolens
Hebeloma sp.
Hemimycena (Mycena) delectabilis
Hemimycena (Mycena) delicatella
Hygrophoropsis (Clitocybe) aurantiaca "False chanterelle"
Hypholoma (Naematoloma) capnoides
Hypholoma (Naematoloma) dispersum
Hypholoma (Naematoloma) fasciculare
Inocybe chelanensis
Inocybe geophylla
Inocybe griseolilacina
Inocybe hirsuta var. maxima
Inocybe lilacina
Inocybe pudica
Inocybe sororia
Laccaria amethysteo-occidentalis (L. amethystina)
Laccaria bicolor
Laccaria laccata
Laccaria sp.
Lactarius deliciosus
Lactarius hepaticus
Lactarius kauffmanii
Lactarius luculentus
Lactarius olympianus
Lactarius rubrilacteus (L. sanguifluus)
Lactarius rufus
Lactarius subflammeus
Lactarius torminosus
Lactarius uvidus
Lactarius sp.
Lepiota rubrotinctoides (L. rubrotincta)
Lepista (Clitocybe) inversa (C. flaccida)
Lepista (Clitocybe) irina
Lepista sp.
Leptonia formosa
Leptonia gracilipes
Leratiomyces ceres (Hypholoma aurantiacum, Stropharia aurantiaca)
Leucoagaricus leucothites (Leucoagaricus naucinus, Lepiota naucina)
Leucoagaricus (Leucoagaricus naucinus, Lepiota naucina)
Leucocoprinus (Lepiota) brebissonii
Lyophyllum decastes
Marasmius copelandii
Marasmius oreades
Melanoleuca melaleuca
Mycena adonis (M. amabilissima)
Mycena amicta
Mycena aurantiidisca
Mycena clavicularis
Mycena elegantula
Mycena epipterygia
Mycena galericulata
Mycena haematopus
Mycena oregonensis
Mycena pura
Mycena purpureofusca
Mycena robusta (M. plumbea)
Mycena rosella
Mycena stipata (M. alcalina)
Mycena strobilinoides
Mycena vulgaris
Mycena spp.
Nolanea bicoloripes
Omphalina sp.
Paxillus involutus
Phaeocollybia kauffmanii
Phaeocollybia sp.
Phaeolepiota (Pholiota) aurea
Pholiota albocrenulata
Pholiota astragalina
Pholiota aurivella
Pholiota decorata
Pholiota flavida
Pholiota limonella
Pholiota malicola
Pholiota malicola var. macropoda
Pholiota (Kuehneromyces) mutabilis
Pholiota squarrosa
Pholiota squarrosoides
Pholiota terrestris
Pleurocybella (Pleurotus) porrigens
Pleurotus pulmonarius (P. ostreatus)
Pluteus cervinus
Pluteus lutescens
Pluteus pellitus
Pluteus sp.
Psathyrella gracilis
Psathyrella hydrophila
Psathyrella sp.
Pseudoarmillariella ectypoides
Rhodocollybia (Collybia) butyracea
Russula aeruginea
Russula brevipes
Russula crassotunicata
Russula dissimulans (R. nigricans)
Russula eleodes
Russula exalbicans
Russula farinipes
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Russula fragilis
Russula murrillii
Russula sanguinaria (R. rosacea)
Russula silvicola
Russula sphagnophila
Russula xerampelina "Woodland Russula", "Shrimp mushroom"
Russula xerampelina var. isabelliniceps
Schizophyllum commune "Splitgill"
Strobilurus (Collybia) trullisatus "Cone lover"
Stropharia aeruginosa
Stropharia ambiguа
Stropharia hormannii
Stropharia rugosoannulata
Tapinella (Paxillus) atrotomentosa
Tricholoma flavovirens
Tricholoma imbricatum
Tricholoma saponaceum
Tricholomopsis decora
Tubaria furfuracea
Xeromphalina campanella
Xeromphalina cornui

Lichens
Cladonia carneola
Cladonia transcendens
Evernia prunastri
Hypogymnia imshaugii
Lichenomphalia (Omphalina) umbellifera (O. ericetorum)
Lobaria oregana
Lobaria pulmonaria
Peltigera neopolydactyla
Ramalina farinacea
Ramalina menziesii
Usnea filipendula
Usnea longissima

Slime molds
Fuligo septica
Lamproderma sp.
Lycogala epidendrum
Stemonitis fusca
Trichia sp.

Species List, Bowman Bay
Foray, November, 2011

GILLED MUSHROOMS

Agaricus augustus
Agaricus campestris
Agaricus comtulus
Agaricus moelleri (A. praeclaresquamosus)
Amanita muscaria “Fly Amanita”
Armillaria sp.
Clitocybe terrestris
Clitocybe fragrans
Clitocybe candidans
Clitocybe sp.
Chlorophyllum brunneum
Chroogomphus tomentosus
Chrysomphalina (Omphalina) aurantiaca)
Clitocybe connata (C. dilatata)
Clitocybe inversa
Collybia sp.
Cortinarius sp. (4)
Gomphidius oregonensis
Gomphidius subroseus
Gymnopilus sapineus
Gymnopilus sp.
Gymnopus dryophilus
Hebeloma crustuliniforme ?
Hygrophoropsis (Clitocybe) aurantiaca “False Chanterelle”
Hypholoma (Naematoloma) fasciculare
Inocybe geophylla
Inocybe lilacina
Inocybe sp.
Laccaria amethysteo-occidentalis (L. amethystina)
Laccaria laccata
Leiota clypeolaria
Leucopaxillus albissimus
Marasmiellus (Marasmius) candidus (M. magnisporus)
Mycena epipterygia var. epipterygia
Mycena pura
Nolanea sericea
Paxillus involutus
Pholiota terrestris

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Pluteus cervinus  
Russula fragilis  
Russula sp.  
Russula xerampelina  
Stropharia ambigua  
Tricholomopsis rutilans  
Tricholoma pessumdatum

NON GILLED MUSHROOMS

Alurea aurantia  
Cantharellula umbonata  
Cantharellus formosus (C. cibarius)

Last Look at the Bizarre Season of 2011 (That Ran Into 2012)  
By Jack Waytz

Just when it seemed that mycorrhizals had taken the year off...

On a warm, wet January 8th, I was walking up the logging road opposite Gate 9 of Sudden Valley, when I
had a feeling I should check one of the mossy berms just a few yards from Lake Louise Road for hedgehog
mushrooms - despite the lateness of the season, and the fact that it had frozen hard several days here.
Over the years, I have learned to trust my “spidy sense” for mushrooms, in, or out, of season. As it turned out, I did not find any hedgehogs, but there were several decent sized clumps of Cantherellus
tubaeiformis (yellowfoot chanterelles) there - in prime condition. This prompted me to set forth the next morning, accompanied by friend and recent addition to our club, Jen Green, on an expedition up the Galbraith Mountain, to an area that I had luck finding the yellowfoot in years past. When we arrived at our destination, I could hardly believe what we found.

In an area no larger than 200 square feet, we gathered enough of these diminutive treats to fill a basket: fully 4.75 pounds worth! At around 15 mushrooms to an ounce, this constitutes several hundred mushrooms! How was so much energy available from the conifer hosts in the dead of winter to support such a massive fruiting of these tasty mushrooms? Yet another myco-mystery to ponder. The next morning, the temperature fell to about 19 degrees, and two days after that, the mountain was under more than two feet of snow. Timing really is everything!